Absolute Stereochemistry of Amphidinolide C

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The absolute configurations at 12 chiral centers in amphidinolide C (1), a potent cytotoxic 25-membered macrolide isolated from a marine dinoflagellate *Amphidinium* sp., were determined to be 3*S*, 4*R*, 6*R*, 7*R*, 8*R*, 12*R*, 13*S*, 16*S*, 20*R*, 23*R*, 24*R*, and 29*S* by combination of NMR analyses, degradation experiments, and synthesis of the C-1–C-7 segment.

Amphidinolides are a series of cytotoxic macrolides possessing unique structural features isolated from laboratorycultured marine dinoflagellates *Amphidinium* sp.¹ Amphidinolides C² (**1**, Scheme 1)) and F,³ isolated from dinoflagellates



Amphidinium sp. (Y-5 and Y-26 strains, respectively), are unique 25-membered macrolides having two tetrahydrofuran rings and vicinally located one-carbon branches. Particularly, amphidinolide C (1) exhibited potent cytotoxicity against tumor cells. The gross structure of 1 has been elucidated by

2D NMR data, and the relative stereochemistry of the C-1– C-8 and C-20–C-23 portions has been assigned tentatively by NOESY correlations of **1** and its 7,8-*O*-isopropylidene derivative (**2**).⁴ During our search for bioactive metabolites from marine dinoflagellates,⁵ relatively large amounts of amphidinolide C (**1**) have been recently isolated from three strains (Y-56,^{1d,e} Y-59, and Y-71) of the genus *Amphidinium*, which were separated from the inside cells of the marine acoel flatworms *Amphiscolops* sp. This sample was utilized to reinvestigate the relative stereochemistry and to determine the absolute configurations at 12 chiral centers in **1**.

Investigation of Relative Stereochemistry. In our previous studies, the relative stereochemistry of H-3/H-4, H-3/

^{(1) (}a) Ishibashi, M.; Kobayashi, J. *Heterocycles* 1997, 44, 543-572.
(b) Tsuda, M.; Endo, T.; Kobayashi, J. *Tetrahedron* 1999, 55, 14565-14570.
(c) Kubota, T.; Tsuda, M.; Kobayashi, J. *Tetrahedron Lett.* 2000, 41, 713-716.
(d) Tsuda, M.; Endo, T.; Kobayashi, J. J. Org. Chem. 2000, 65, 1349-1352.
(e) Kobayashi, J.; Kubota, T.; Endo, T.; Tsuda, M. J. Org. Chem. 2001, 66, 134-142.

⁽²⁾ Kobayashi, J.; Ishibashi, M.; Wälchli, M. R.; Nakamura, H.; Hirata, Y.; Sasaki, T.; Ohizumi, Y. J. Am. Chem. Soc. **1988**, 110, 490–494.

⁽³⁾ Kobayashi, J.; Tsuda, M.; Ishibashi, M.; Shigemori, H.; Yamasu, T.; Hirota, H.; Sasaki, T. J. Antibiot. **1991**, 44, 1259-1261.

⁽⁴⁾ Ishiyama, H.; Ishibashi, M.; Kobayashi, J. Chem. Pharm. Bull. 1996, 44, 1819–1822.

⁽⁵⁾ Kobayashi, J.; Shimbo, K.; Sato, M.; Shiro, M.; Tsuda, M. Org. Lett. **2000**, *2*, 2805–2807.

H-6, and H-20/H-23 on the two tetrahydrofuran rings was proposed to be all *anti* from NOESY data of amphidinolide C (1).⁶ An *erythro* relationship for the 7,8-diol was deduced from analysis of the NOESY spectrum of the 7,8-*O*-isopropylidene derivative (2) of 1. The ${}^{3}J$ (H-12,H-13) (8.8 Hz) was a typical value for an *anti* relationship⁷ (Figure 1a).



Figure 1. Rotation models for (a) C-12–C-13 and (b) C-23–C-24 bonds of amphidinolide C (1). NOESY correlations are illustrated by solid arrows.

The values for ${}^{2}J(C-13,H-12)$ (-6.3 Hz), ${}^{3}J(C-14,H-12)$ (3.1 Hz), and ${}^{3}J(C-38,H-13)$ (3.5 Hz), which were obtained from the hetero half-filtered TOCSY (HETLOC)⁸ spectrum, indicated that H-12 was gauche to 13-OH, while H-12 and H-13 were gauche to C-14 and C-38, respectively. The gauche relation between the C-13-C-14 and C-12-C-38 bonds was deduced from the intense NOESY correlation for H-14 ($\delta_{\rm H}$ 2.53)/H₃-38. Thus, the *erythro* relation for the C-12-C-13 bond was established. On the other hand, an anti relationship for H-23 and H-24 (Figure 1b) was inferred from the ${}^{3}J(H-23,H-24)$ value (7.7 Hz). The NOESY correlation for H-22 ($\delta_{\rm H}$ 1.60)/H-24 as well as the J(C,H) values for C-23/H-24 (-4.4 Hz) and C-22/H-24 (1.4 Hz) indicated that C-22 and 23-O were both gauche to H-24. The NOESY cross-peak for H-22 ($\delta_{\rm H}$ 1.87)/H-25 was suggestive of the gauche relation between C-22-C-23 and C-24-C-25 bonds, and the ${}^{3}J(C-25,H-23)$ value (1.0 Hz) was a typical one for a gauche relation, thus indicating that the relative configuration of C-23-C-24 was three.

Absolute Configurations at C-13 and C-29. Determination of the absolute configurations of two oxymethine carbons at C-13 and C-29 was accomplished by a modified Mosher method.⁹ The 7,8-*O*-isopropylidene derivative (2) of amphidinolide C (1) was treated with (*R*)-(-)- and (*S*)-(+)-2-methoxy-2-trifluoromethyl-2-phenylacetyl chloride (MT-PACl) to afford the bis-(*S*)- and bis-(*R*)-MTPA esters (**3a** and **3b**, respectively), respectively. $\Delta\delta$ values ($\delta_S - \delta_R$) are shown in Figure 2. The $\Delta\delta$ values for H₂-14, H₂-17, H₂-31, H₂-32, H₂-33, H₃-34, H₃-39, and H₂-41 were negative, while



Figure 2. $\Delta\delta$ values [$\Delta\delta$ (in ppm) = $\delta_S - \delta_R$] obtained for the (*S*)- and (*R*)-MTPA esters (**3a** and **3b**, respectively) of the 7,8-*O*-isopropylidene derivative (**2**) of amphidinolide C (**1**).

positive $\Delta\delta$ values were observed for H-7, H-8, H-10, H-12, H-23, H-24, H-25, H-26, H-27, H₂-36, H₃-37, H₃-38, and H₃-40, thus indicating that C-13 and C-29 both had *S*-configurations.

Absolute Configurations at C-3, C-4, and C-6. To investigate the absolute stereochemistry at C-3, C-4, and C-6, the oxidative degradation reaction for the 7,8-diol unit in amphidinolide C (1) was performed as follows. Reduction of 1 with DIBAL, oxidative cleavage of the 7,8-diol unit with NaIO₄, reduction with NaBH₄, esterification with (R)-(-)-MTPACl, and then HPLC separation furnished the bis-(S)-MTPA ester (4a) of the C-1–C-7 segment (Scheme 2),



of which the structure was elucidated by analysis of ${}^{1}\text{H}{-}{}^{1}\text{H}$ COSY and NOESY spectra. On the other hand, both bis-(*S*)- and -(*R*)-MTPA esters (**4a** and **4b**, respectively) of the C-1–C-7 segment were prepared from the (4*R*,6*R*)-6hydroxymethyl-4-methyl- γ -butyrolactone (**7**), which was derived from D-glutamic acid¹⁰ (Scheme 3). Two-carbon



⁽⁶⁾ Kobayashi, J.; Ishibashi, M. Chem. Rev. 1993, 93, 1753-1769.

 ⁽⁷⁾ Matsumori, N.; Kaneno, D.; Murata, M.; Nakamura, H.; Tachibana,
 K. J. Org. Chem. 1999, 64, 866–876.

^{(8) (}a) Otting, G.; Wüthrich, K. Q. Rev. Biophys. **1990**, 23, 39–96. (b) Wollborn, U.; Leibfritz, D. J. Magn. Reson. **1992**, 98, 142–146. (c) Kurz, M.; Schmieder, P.; Kessler, H. Angew. Chem., Int. Ed. Engl. **1991**, 30, 1329–1331.

⁽⁹⁾ Ohtani, I.; Kusumi, T.; Kashman, Y.; Kakisawa, H. J. Am. Chem. Soc. 1991, 113, 4092–4095.

elongation of the γ -butyrolactone (7) using a Wittig reaction gave a *E*-olefin **8** in 90% yield in two steps. The unsaturated ester **8** was converted into a tetrahydrofuran (94%) by treatment with *tert*-butylammonium fluolide (TBAF) in THF through diastereoselective Michael reaction, and then the ester carbonyl group was reduced by DIBAL to give compound **9** in 91% yield. Relative configurations of H-3/ H-4 and H-3/H-6 in **9** were both assigned as *anti* by NOESY correlations for H₂-2/H-6 and H-3/H₃-35. Deprotection of the benzyl group in **9** was achieved by hydrogenation using palladium—charcoal in EtOH, and then esterification with (*R*)-(-)- and (*S*)-(+)-MTPACl afforded the bis-(*S*)- and (*R*)-MTPA esters (**4a** and **4b**, respectively) of the C-1-C-7 segment.

The ¹H NMR spectrum of the bis-(*S*)-MTPA ester (**4a**) of the C-1–C-7 segment obtained from natural amphidinolide C (**1**) was compared with those of synthetic bis-(*S*)- and -(*R*)-MTPA esters (**4a** and **4b**) of the C-1–C-7 segment (Figure 3). Though **4a** and **4b** showed very similar NMR profiles,



Figure 3. ¹H NMR spectra (partial) of (a) bis-(S)-MTPA ester (**4a**) of the C-1–C-7 segment derived from amphidinolide C (**1**) and (b) synthetic bis-(S)- and (c) bis-(R)-MTPA esters (**4a** and **4b**, respectively) of the C-1–C-7 segment.

significant differences were observed for signals due to the methylene proton at C-7 (**4a**, $\delta_{\rm H}$ 4.06 and 3.98; **4b**, $\delta_{\rm H}$ 4.13 and 3.79). ¹H NMR data of the bis-(*S*)-MTPA ester (**4a**)

derived from a natural specimen were identical with those of the synthetic bis-(S)-MTPA ester (4a). Therefore, the absolute configurations at C-3, C-4, and C-6 were determined to be *S*, *R*, and *R*, respectively.

Absolute Configurations at C-7, C-8, and C-24. The absolute configuration at C-24 was elucidated by application of a modified Mosher method⁸ to the linear methyl esters of amphidinolide C (1). Treatment of amphidinolide C (1) with K_2CO_3 in MeOH yielded a mixture of four linear methyl esters generated by epimerization of C-16 and C-20.¹¹ One of the four methyl esters purified by C₁₈ HPLC was treated with (*R*)-(-)- and (*S*)-(+)-MTPACl to afford the pentakis-(*S*)-(-)- and -(*R*)-(+)-MTPA esters (**5a** and **5b**, respectively). $\Delta\delta$ values obtained from the ¹H chemical shifts of **5a** and **5b** are shown in Figure 4. The $\Delta\delta$ values of the protons from C-19 to C-22 are negative in sign, while those of H-25, H-26, and H-27 are positive, suggesting a 24*R* configuration.

The absolute stereochemistry of C-7 and C-8 was elucidated on the basis of the application of Mosher's method for *erythro*-glycol proposed by Kusumi et al.¹² Negative $\Delta\delta$ values for H₂-2, H-3, H-4, H₂-5, and H-6 and positive ones for H-10, H₂-36, and H₃-37 obtained from **5a** and **5b** indicated that **5a** and **5b** had the 7*S* and 8*R* configurations. Therefore, the absolute configurations at C-7 and C-8 were concluded to be both *R*.

Absolute Configuration at C-16. To determine the absolute configuration at C-16 of amphidinolide C (1),



Baeyer–Villiger oxidation using trifluoroperacetic acid¹³ (TFPA) was applied to obtain the segment including the



Figure 4. $\Delta\delta$ values [$\Delta\delta$ (in ppm) = $\delta_S - \delta_R$] obtained for the pentakis-(*S*)- and -(*R*)-MTPA esters (**5a** and **5b**, respectively) of the linear methyl ester of amphidinolide C (1).

methine carbon at C-16. Amphidinolide C (1) was treated with TFPA followed by reduction with LiAlH₄, esterification with (*S*)-(+)-MTPACl, and HPLC separation to afford a bis-(*R*)-MTPA ester (**6a**) of 1,3-butanediol corresponding to the C-16–C-18 segment of 1 (Scheme 4). On the other hand, the two authentic bis-(*R*)-MTPA esters of (*S*)-(+)- and (*R*)-(-)-1,3-butanediols (**6a** and **6b**, respectively) were prepared. ¹H NMR data of compounds **6a** from a natural specimen were identical with those of synthetic 16*S*-isomer (Figure 5), indicating that the absolute configuration at C-16 of



Figure 5. ¹H NMR spectra (partial) of (a) bis-(R)-MTPA ester (**6a**) of the C-16–C-18 segment derived from amphidinolide C (**1**), (b) the bis-(R)-MTPA ester (**6a**) of (S)-1,3-butanediol, and (c) the bis-(R)-MTPA ester (**6b**) of (R)-1,3-butanediol.

amphidinolide C (1) was determined to be *S*. Therefore, the absolute configurations of 12 chiral centers in amphidinolide

C (1) were elucidated to be 3*S*, 4*R*, 6*R*, 7*R*, 8*R*, 12*R*, 13*S*, 16*S*, 20*R*, 23*R*, 24*R*, and 29*S*.

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Supporting Information Available: Experimental procedures, spectral data of 1, 2, 3a, 3b, 4a (natural and synthetic), 4b, 5a, 5b, 6a (natural and synthetic), and 6b, and Tables S1 and S2. This material is available via the Internet at http://pubs.acs.org.

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^{(10) (}a) Tomioka, K.; Cho, Y.-S.; Sato, F.; Koga, K. J. Org. Chem. **1988**, 53, 4094–4098. (b) Nishida, Y.; Konno, M.; Fukushima, Y.; Ohrui, H.; Meguro, H. Agric. Biol. Chem. **1986**, 50, 191–193.

⁽¹¹⁾ Two of the four linear methyl esters were suggested to have a syn relationship for H-20-H-23 by NOESY correlation for H-20/H-23, while the relative stereochemistry of H-20-H-23 of the two other diastereomers was anti. Epimerization at Č-20 is explained by inversion of C-20, generated through retro-Michael-type cleavage between the ether oxygen and C-20 followed by Michael-type re-formation of the ether linkage. Although such a retro-Michael-Michael reaction for another tetrahydrofuran ring (C-3-C-6) might occur, the anti relationship of H-3/H-4 in the tetrahydrofuran ring was reported to be generally more kinetically and thermodynamically stable than a syn relationship. Methanolysis of 1 with K_2CO_3 in MeOH- d_4 afforded four linear methyl esters labeled with deuterium. In the ¹H NMR data of four linear methyl esters, the signals of α-protons (H2-2, H2-14, H-16, H₂-17, and H₂-19) of three carbonyl groups at C-1, C-15, and C-18 disappeared. This indicated that one of the two 20,23-syn linear compounds and one of two 20,23-anti compounds were products due to epimerization at C-16. Nevertheless, the stereochemisry at C-16 of each compound was not determined. The diastereomer using the modified Mosher method possessed a 20,23-anti relationship.

⁽¹²⁾ Ichikawa, A.; Takahashi, H.; Ooi, T.; Kusumi, T. *Biosci. Biotechnol. Biochem.* **1997**, *61*, 881–883.

⁽¹³⁾ Bailey, A. S.; Gilpin, M. L. Jones, E. R. H. J. Chem. Soc., Perkin Trans. 1 1977, 265–270.